

Comparative analyses of response surface methodology and artificial neural networks on incorporating tetracaine into liposomes

Ana Karina Vidal Pereira^{1#}, Raquel de Melo Barbosa^{2,3,5*#}, Marcelo Augusto Costa Fernandes^{4,6},
Leandro Finkler¹, Christine Lamenha Luna Finkler¹

¹Federal University of Pernambuco, Academic Centre of Vitória, Vitória de Santo Antão, PE, Brazil, ²Department of Pharmacy, UNINASSAU College, Natal, RN, Brazil, ³Department of Pharmacy, Federal University of Rio Grande do Norte, Natal, RN, Brazil, ⁴Department of Computer Engineering and Automation, Federal University of Rio Grande do Norte, Natal, RN, Brazil, ⁵Current affiliation: MIT Department of Chemical Engineering, Massachusetts Institute of Technology, Cambridge, MA, USA, ⁶John A. Paulson School of Engineering and Applied Sciences, Harvard University, Cambridge, MA, USA

This study evaluated the incorporation of tetracaine into liposomes by RSM (Response Surface Methodology) and ANN (Artificial Neural Networks) based models. RCCD (rotational central composite design) and ANN were performed to optimize the sonication conditions of particles containing 100 % lipid. Laser light scattering was used to perform measure hydrodynamic radius and size distribution of vesicles. The liposomal formulations were analyzed by incorporating the drug into the hydrophilic phase or the lipophilic phase. RCCD and ANN were conducted, having the lipid/cholesterol ratio and concentration of tetracaine as variables investigated and, the encapsulation efficiency and mean diameter of the vesicles as response variables. The optimum sonication condition set at a power of 16 kHz and 3 minutes, resulting in sizes smaller than 800 nm. Maximum encapsulation efficiency (39.7 %) was obtained in the hydrophilic phase to a tetracaine concentration of 8.37 mg/mL and 79.5:20.5% lipid/cholesterol ratio. Liposomes were stable for about 30 days (at 4 °C), and the drug encapsulation efficiency was higher in the hydrophilic phase. The experimental results of RCCD-RSM and ANN techniques show ANN obtained more refined prediction errors than RCCD-RSM technique, therefore, ANN can be considered as an efficient mathematical method to characterize the incorporation of tetracaine into liposomes.

Keywords: Local anesthetics. Liposomes. Response Surface Methodology. Artificial Neural Networks.

INTRODUCTION

The mathematical modeling of drugs encapsulation has been a research target in the recent years. Based in the mathematical modeling, the researchers can perform estimates about the parameters and minimize the laboratory experiments. There are many techniques can be used for mathematical modeling of drugs encapsulation (Siepmann, Siepmann, 2008). However, methods based on the Response Surface Methodology (RSM) and Artificial Neural Networks (ANN) have been showing satisfactory results (Sun *et al.*, 2003).

The use of statistical experimental planning and RSM explores the relationships between several explanatory variables and one or more response variables

and is one of the most popular methods in the optimization of drug delivery systems. The optimization of variables by RSM includes statistical experimental designs, multiple regression analysis, and mathematical optimization algorithms, used to select the optimum formulation (Subramanian, Yajnik, Murthy, 2004). However, in general, the predictions of response variables are done by a second-order equation. Since this prediction is often limited to low levels, ANN was incorporated to overcome this limitation (Takayama, Fujikawa, Nagai, 1991; Sulaiman *et al.*, 2017; Moussa *et al.* 2017).

Prediction techniques based on Artificial Intelligence (AI) has been used to characterize formulations and biotechnological processes. Among the AI techniques can highlight the experimental prediction models represented by ANN (Takayama *et al.*, 2003; Subramanian, Yajnik, Murthy, 2004; Zhang *et al.*, 2010). The ANN is generic mathematical models inspired by biological neurons, and other actions work with interpolation (or predictors) universal. These

*Correspondence: R. M. Barbosa. Departamento de Farmácia, UNINASSAU. Avenida Engenheiro Roberto Freire 1514, Capim Macio, ZP 59082-095, Natal, RN – Brazil. Phone: 55 84 99612-4869. E-mail: m.g.barbosafernandes@gmail.com.
#These authors contributed equally to this work.

structures can learn and can help without any experimental model from a set of data called training data (Haykin, 1998).

Recent researches have been published comparing the data analysis techniques RCCD- RSM and ANN (Desai *et al.*, 2008; Asiltürk, Çunkaş, 2011; Moghaddam, Khajeh, 2011; Ahmadi, Golian, 2011; Maran, Priya, 2015). According to the most of articles presented, the ANN is an alternative better than the methods based on RSM in the case of performance. Furthermore, ANN can increase the level of certainty associated with the results and simultaneously can be used to validation new technological strategies.

The liposomes containing local anesthetic tetracaine, and carriers were prepared in two stages: a hydrophilic phase by incorporating the drug during hydration of the lipid film and a lipophilic phase, incorporating tetracaine during the film preparation. The two forms of preparation were carried out to determine which mathematical modeling approach could result in greater drug encapsulation. In both, the sonication parameters (frequency and duration) were fixed, and the components were changed.

The experiments were performed using an experimental design since there is a substantial lack of application of this statistical tool in research involving systems of controlled drug release. In both cases, RCCD experimental designs and ANN were conducted, having as variables investigated the lipid/cholesterol ratio and the concentration of tetracaine and, encapsulation efficiency and the average size of liposomes as response variables.

The objective of this study was to analyze two different approaches (RSM and ANN) of the mathematical modeling applied to the incorporating tetracaine into liposomes and compare their results. There have been many works in the literature about the encapsulating tetracaine in liposomes (Friedmann, Juliano, 1984; Foldvari, Jarvis, Oguejiofor, 1993; Fisher *et al.*, 1994; El Maghraby, Arafa, Essa, 2018). However, there have been fewer studies about the different mathematical modeling approaches and comparative analyzes. The mathematical modeling studies are necessary to understand the incorporation process and show the optimization alternatives to the process.

MATERIAL AND METHODS

Material

Liposomes containing tetracaine (TCC) were prepared using the following reagents: hydrogenated soy phosphatidylcholine (via Farma®, São Paulo, Brazil), tetracaine (Sigma-Aldrich Co., St. Louis®, MO, USA), chloroform (Merk®, Darmstadt, Germany), methanol

(Merk®, Darmstadt, Germany) and cholesterol (Sigma-Aldrich Co., St. Louis®, MO, USA). All other chemicals were of analytical grade.

Methods

Preparation of liposomes

The preparation of liposomes was performed by the method of Bangham (New, 1990) by rehydrating the dried film with the subsequent homogenization of vesicles size (Lasic, 1993). The phospholipid stored at -20 °C was weighed according to the desired composition and then solubilized with cholesterol in 10 mL of a 9:1 chloroform/methanol solution (v/v). The solution was homogenized for 5 minutes in a rotary evaporator, followed by solvent evaporation under vacuum (650 mmHg) at a temperature above the lipid phase transition ($T_t = 33$ °C) until the dry film formation. In the step of dry film hydration was added 10 mL of a 60:40 ethanol/water solution (% v/v) at room temperature.

Initially, liposomes containing 100% lipid (0.05 g lipid/10 mL solvent) were added in order to optimize the sonication conditions. This was evaluated from a RCCD experimental design (rotational central composite design) with repetition in triplicate at the central point (level 0) and four axial points (levels $\pm \alpha$, where $\alpha = 1.41$), totaling 11 trials. The independent variables were time and sonication power and the dependent variable (response) was liposome size. The experiments were performed randomly and TABLE I show the coded levels and actual values of the variables.

After defining the optimal conditions of sonication, liposomes containing tetracaine were prepared according to two experimental designs. At first, the drug was incorporated during the lipid film hydration, corresponding to the hydrophilic phase. In the second experimental design, liposomes were prepared with the drug incorporation during the lipid film preparation (drug in the lipophilic phase). In both cases, a RCCD experimental design was performed, having the lipid-cholesterol ratio and drug concentration as independent variables and the liposome size and encapsulation efficiency as dependent variables. The hydration solution was composed of a 60:40 ethanol/water solution (% v/v) for solubilizing the tetracaine at room temperature. The Table II shows the coded levels and actual values of the variables studied.

Homogenization of vesicles size

The samples obtained by the dry film hydration method were kept at rest for 30 minutes to reach

TABLE I - Coded levels and actual values of the variables time and sonication power

Assays	Time		Power	
	Coded	Value (min)	Coded level	Value (kHz)
1	-1.00	1.58	-1.00	11.4
2	+1.00	4.42	-1.00	11.4
3	-1.00	1.58	+1.00	18.4
4	+1.00	4.42	+1.00	18.4
5	-1.41	1.00	0.00	14.9
6	+1.41	5.00	0.00	14.9
7	0.00	3.00	-1.41	10.0
8	0.00	3.00	+1.41	19.8
9	0.00	3.00	0.00	14.9
10	0.00	3.00	0.00	14.9
11	0.00	3.00	0.00	14.9

TABLE II - Coded levels and actual values of the variables concentration of lipids: cholesterol and tetracaine concentration to the experiments performed in both hydrophilic and lipophilic phases

Assays	Lipid:cholesterol ratio		Tetracaine	
	Coded level	Value (m%/m%)*	Coded level	Value (mg/mL)
1	-1.00	57:43	-1.00	2.5
2	-1.00	57:43	+1.00	8.5
3	+1.00	93:7	-1.00	2.5
4	+1.00	93:7	+1.00	8.5
5	0.00	75:25	-1.41	1.0
6	0.00	75:25	+1.41	10.0
7	-1.41	50:50	0.00	5.5
8	+1.41	100:0	0.00	5.5
9	0.00	75:25	0.00	5.5
10	0.00	75:25	0.00	5.5
11	0.00	75:25	0.00	5.5

* 100% corresponds to a mass of 0.05 g in 10 mL of solvent

equilibrium before vesicles size homogenization. The liposome preparation was performed by sonication method using an ultrasonic sonicator (Unique®) in the optimized conditions obtained in the experimental design (3 min and 16 KHz).

Determination of the hydrodynamic radius and vesicles size distribution

The hydrodynamic radius and liposomes size distribution were determined by the dynamic light scattering (DLS) using a laser (Malvern Zetasizer Nanoseries/NANO ZS90) in 1 mL of dispersion at 25 °C.

Encapsulation efficiency (EE%)

Encapsulation efficiency was determined by ultracentrifugation (refrigerated centrifuge/Hermle Z36HK). The samples were centrifuged at 30,000 rpm for 2 h at 10 °C and the supernatants diluted in 100 mL of a 60:40 ethanol/water (% v/v) to determine the concentration of non-encapsulated tetracaine. This technique was chosen based on the description in the literature that there is no difference between the method of dialysis through membranes and the ultracentrifugation method (López-Pinto, González-Rodríguez, Rabasco, 2005; Nasr *et al.*, 2008). The encapsulation efficiency (EE%) was determined by Equation 1:

$$EE\% = \left[\frac{\text{Total Conc. TTC} - \text{non-encapsulated Conc. TTC}}{\text{Total Conc. TTC}} \right] \times 100 \quad (1)$$

where *Total Conc. TTC* is the initial concentration of tetracaine and non-encapsulated *Conc. TTC* is the tetracaine concentration in the supernatant.

Determination of tetracaine concentration

Initially, solutions containing tetracaine were prepared in 60:40 ethanol/water solution (% v/v) at different ranges of concentration (0 to 0.001 mg/mL and 0 to 0.015 mg/mL, the latter being the maximum concentration that allowed a linear fit between the variables). From the experimental data were plotted standard curves of absorbance (analyzed by UV spectrophotometry at 310 nm) versus concentration. All experiments were carried out in five repetitions. The tetracaine concentrations were determined in the supernatant after centrifugation (non-encapsulated drug).

Stability study

Stability study was performed with tetracaine-loaded liposomes stored at 4 °C over 15 days. Samples were taken at 0, 7 and 15 days for measurement of drug loading as described at Encapsulation efficiency. Tetracaine content remaining in the liposome formulations were assessed by spectrophotometry as described in Determination of tetracaine concentration (Badran, Shalaby, Al-Omrani, 2012)

Prediction models based on RCCD

A central composite design (CCD) is an experimental design for building a second order (quadratic) model for the response variable. The CCD contains an imbedded

factorial or fractional factorial design with center points that is augmented with a group of axial points ('star points') that allow estimation of curvature. If the distance from the center of the design space to a factorial point is ± 1 unit for each factor, the distance from the center of the design space to a star point is $|\alpha| > 1$. The factorial points contribute to the estimation of linear terms and interactions between factors and the center points provide information on the existence of curvature in the system. If the curve is found, the addition of the axial points allows to efficiently estimate the quadratic terms. In the rotatable CCD (RCCD), the $|\alpha|$ is 1.41 for two variables and describe a circular design geometry (Montgomery, 1997).

Prediction models based on ANN

The Multi-Layer Perceptron (MLP) trained with the Levenberg-Marquardt backpropagation algorithm is a technique that is widely used in applications based on artificial neural networks (Haykin, 1998; Levenberg, 1944; Marquardt, 1963). Figure 1 shows a three-layer MLP type network with P neurons in the input layer, K neurons in the hidden layer, and H neurons in the output layer. The i -th neuron of the hidden layer processes the signal according to Equation 2:

$$z_i = \phi \left(\sum_{j=0}^{P-1} w_{ij}^1 x_j \right) \quad (2)$$

where z_i ($i = 0, 1, \dots, K-1$) is the output signal of the i -th neuron, u_j ($j = 0, 1, \dots, P-1$) is the j -th input signal, w_{ij}^1 are the weights associated with the connections between the input, x_j , and the output, z_i , and $\phi(\cdot)$ is the activation function of the hidden layer. The output of the h -th neuron of the output layer is given by Equation 3:

$$y_h = \phi \left(\sum_{i=0}^{K-1} w_{hi}^2 z_i \right) \quad (3)$$

where y_h ($h = 0, 1, \dots, H-1$) is the output signal of the h -th neuron, w_{hi}^2 are the weights associated with the connections between the output of the hidden layer, z_i , and the output of the output layer, y_h , and $\phi(\cdot)$ is the activation function of the output layer.

Thus, to improve the results of the analysis, an ANN type MLP trained with Levenberg Marquardt backpropagation algorithm was used. All experiments used two inputs (x_1 and x_2), N neurons in the hidden layer, and one neuron in the output layer. The input and output data were normalized between zero and one in order to balance the training process (Zhang *et al.*, 2010).

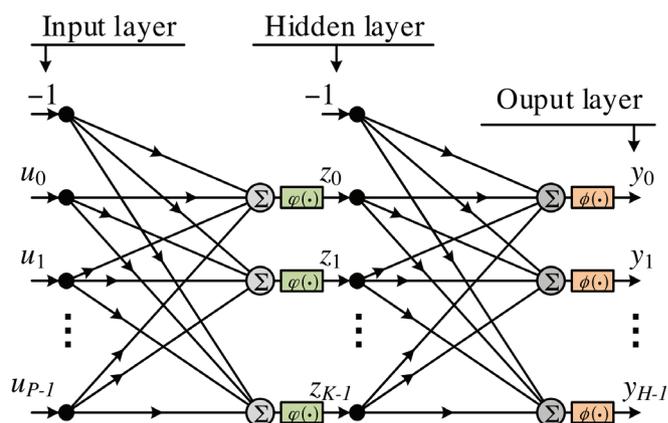


FIGURE 1 - The Multi-Layer perceptron with three-layer. The network with P neurons in the input layer, K neurons in the hidden layer, and H neurons in the output layer.

RESULTS AND DISCUSSION

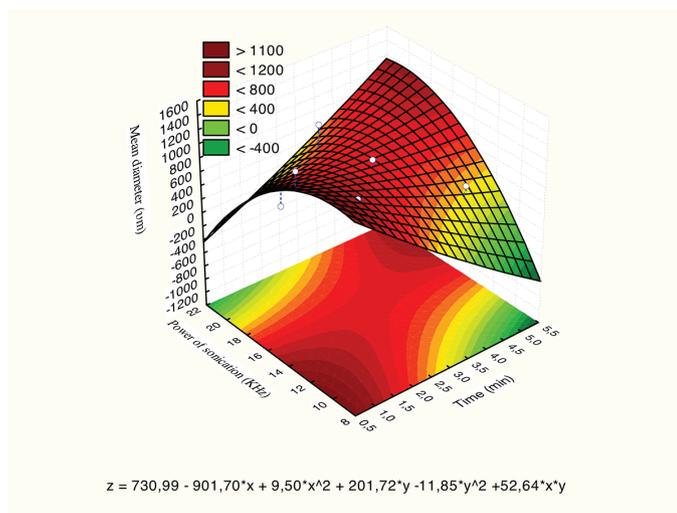
Experimental design of sonication

Figure 2 shows the response surface (A) and contour curve (B) for the variable mean diameter of liposomes containing 100% lipid (0.05 g/10 mL). The values of power and sonication time corresponding to 16 kHz and 3 min, respectively, produce liposomes with a size smaller than 800 nm. Thus, these sonication conditions were selected for further experiments due to decreasing the particles size that produced and desire.

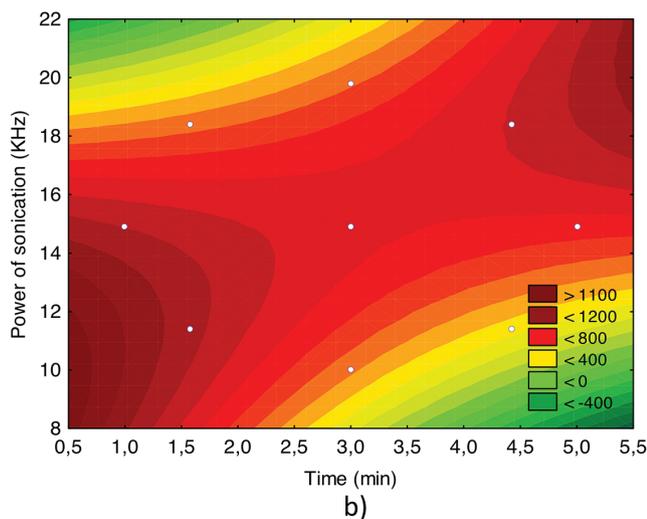
The liposomes obtained in the LUV diameter were chosen for being able to extravasate blood into the interstitial space without causing pulmonary embolism (Lasic, 1993; Sapa, Allen, 2003). Furthermore, studies on the vesicles size show that the rate of recognition and removal by phagocytic defense system varies according to the diameter with a long half-life of liposomes (> 1000 nm) of 0.2 h while the shortest have a 1.5 h half-life (Laverman *et al.*, 1999).

Parameters as the power above 16 kHz and times of sonication longer than 4.5 min during producing of liposomes led to obtaining samples with loss of physical stability, being observed the formation of viscous foam in suspension, resulting in a subsequent increase in vesicles diameter.

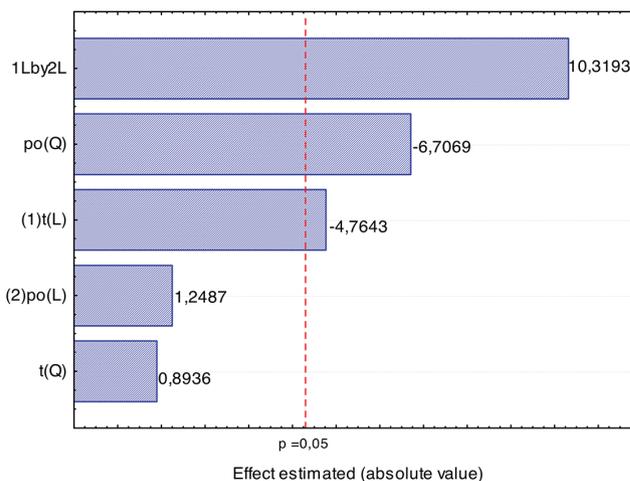
Figure 2C shows the Pareto diagram, in which we can observe that the quadratic term of the variable power of sonication and linear terms of the variables time and power \times time interaction were significant. The empirical model that describes the behavior of variables in the experimental conditions investigated is shown in Equation 4, with the significant terms in bold, where D is the diameter, t is time and P power of sonication.



a)



b)



c)

FIGURE 2 - Response surface (A), contour curve (B) and Pareto chart (C) for the mean diameter of liposomes depending on the power and sonication time (liposomes containing 100% - 0.05 g/10 mL soy phosphatidylcholine phospholipid).

$$D = 730.99 - 901.70*t + 9.50*t^2 + 201.72*P - 11.85*P^2 + 52.64*t*P \quad (4)$$

The effect of tetracaine-loaded on liposome stability

The drug encapsulation over 15 days was performed in the liposomal formulations to verify the integrity of the vesicles, considering that was used a hydration solution of 60:40 ethanol/water solution (% , v/v). The stability assays show that was not observed drug loss from the liposomes for the different formulations at 4 °C after 15 days of storage (data not shown). Ethanol at 40%, v/v was used to dissolve tetracaine, on the other hand, could also dissolve phospholipid and cholesterol causing rupture of the liposomes, but it was not observed. Besides this, the

presence of ethanol increases the permeability for topical application. According to Roberts and Water (Roberts, Walker, 1993), the use of 60 % water increase hydration of the stratum corneum and enhances transdermal flux of a variety of drugs.

Mura and collaborators (2007) studied vesicles entrapping benzocaine prepared using 50:50 w/w phosphatidylcholine-cholesterol as the lipophilic phase and 50:50 v/v ethanol-water as the hydrophilic phase. Similarly, to this work, the addition of the drug into the hydrophilic phase enabled an improvement of entrapment efficiency.

Experimental design - liposomes prepared in hydrophilic phase

Figure 3A and 3B show, respectively, the response

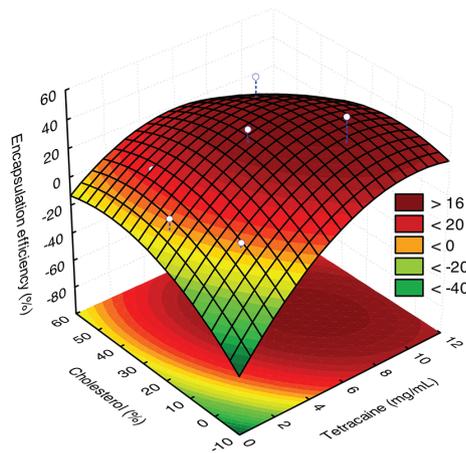
surface and the contour curve when the variable studied was the encapsulation efficiency of liposomes prepared in the hydrophilic phase. The optimal region for encapsulation efficiency was achieved. The critical values of the model that maximize the response were equivalent to a tetracaine concentration of 8.37 mg/mL and 79.5:20.5 lipid/cholesterol ratio (% w/w) for 39.7 % maximum encapsulation efficiency.

Figure 3C shows, Pareto chart, that the only significant effect ($p < 0.05$) is the linear term of the variable tetracaine. This term is in bold in Equation 5, where E represents the encapsulation efficiency, the TTC tetracaine concentration, and Co cholesterol content.

$$E = -41.51 + 15.78 * TTC - 0.83 * TTC^2 + 1.48 * Co - 0.02 * Co^2 - 0.09 * TTC * Co \quad (5)$$

In order to confirm the results predicted by the model, experiments were performed in triplicate under the optimum condition expected (8.37 mg/mL and 79.5:20.5 lipid/cholesterol ratio (% w/w)). Results are shown in TABLE II, yielding a mean value of $32.3 \pm 7.5 \%$, close to the value predicted by the model (39.7 %).

For being amphiphilic molecules, local anesthetics have a high affinity for cell membrane (Covino, 1986). The theories proposed to explain the mechanisms of action of these compounds can be classified into two categories. One that attributes the anesthetic effect to the binding on the voltage-dependent axons sodium channel protein. Another mechanism of action considers the interaction of local anesthetics with membrane lipids (known as the "lipid hypothesis") being responsible for the changes in sodium channel protein, and therefore to blockage in



$$z = -41,51 + 15,78 * x - 0,83 * x^2 + 1,48 * y - 0,02 * y^2 - 0,09 * x * y$$

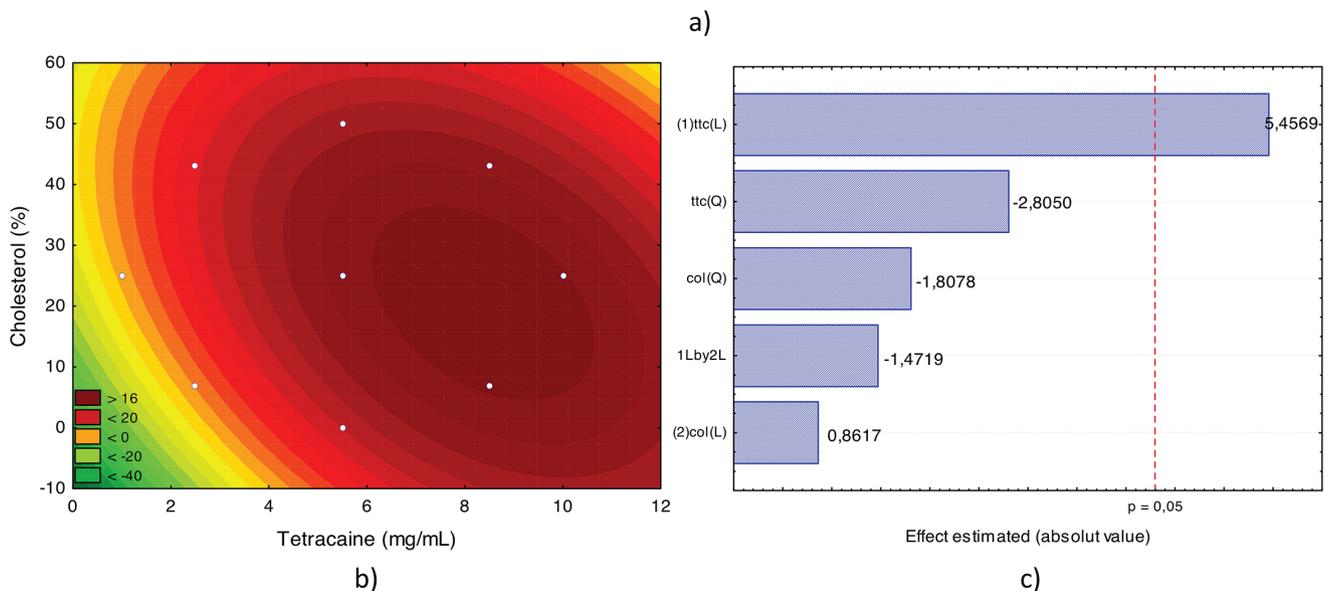


FIGURE 3 - Response surface (A), contour curve (B) and Pareto chart (C) for the encapsulation efficiency of liposomes prepared in the hydrophilic phase.

TABLE III - Results of experiments performed in triplicate using 79.5:20.5 lipid: cholesterol ratio (% , w/w) and tetracaine concentration of 8.37 mg / mL for liposomes prepared in hydrophilic phase

Assay	Lipid:cholesterol ratio	Tetracaine	E (%)**
	Value (m%/m%)*	Value (mg/mL)	
1	79.5:20.5	8.37	32 ± 3
2	79.5:20.5	8.37	25 ± 5
3	79.5:20.5	8.37	40 ± 4

* 100% corresponds to a mass of 0.05 g in 10 mL of solvent. **Maximum encapsulation efficiency predicted by model: 39.7 %

the nervous stimulus conduction. Despite these theories, studies have reported that tetracaine incorporated into phosphatidylcholine, phosphatidylserine and biological membranes was expelled from the bilayer by increasing pressure (Auger *et al.*, 1987), an observation confirmed by the infrared technique.

The literature provides reports of the action of local anesthetics on the decrease of lipid phase transition temperature (Tt) studied by different techniques. Studies using differential scanning calorimetry registered a Tt decrease of the phosphatidylcholine vesicles caused by the partition of benzocaine, procaine, tetracaine, lidocaine and dibucaine in concentrations equal to those required for 50% block in the nervous stimulus conduction (Lee, 1978).

De Paula and collaborators have studied the interaction of local anesthetics with lipid membranes using NMR, EPR and fluorescence techniques (de Paula, Schreier, 1996; Fraceto, de Paula, 2006). For amino-amide local anesthetics (lidocaine, mepivacaine, bupivacaine, and etidocaine) was demonstrated the differential effect of local anesthetics on the bilayer regions monitored by different paramagnetic probes. Besides, changes in lipids longitudinal relaxation times in the presence of local anesthetics and specific intermolecular interactions between the hydrogen of local anesthetics and liposomes provided evidence about their location within the bilayer (Fraceto, de Paula, 2006).

A fact that should be taken into consideration is that phosphate groups have negative charges, which can stabilize positive charges of local anesthetics. There are several reports in the literature that showed of local anesthetics competition with calcium ions by interaction with phosphate groups of lipid vesicles (de Paula, Schreier, 1996; Cullis, de Kruijff, 1978). This is the preferred pathway, i.e., the place where local anesthetics should meet most of the time. However, these amphiphilic molecules are spread quickly across the biological membrane and the aqueous phase (de Paula, Schreier, 1996) or are in dynamic balance, with preferential insertion within the bilayers. This location, determined by the local anesthetic

interactions with phospholipids must be decisive for its the specific action (Ragsdale *et al.*, 1994).

Figure 4A and 4B show, respectively, the response surface and contour curve for analysis of the mean diameter of liposomes. Liposomes type LUV with size <1000 nm, suitable for drug delivery system, were obtained for samples with 79.5:20.5 lipid: cholesterol ratio (% , w/w) and the 8.37 mg/ml tetracaine concentration (optimal values according to Figure 4). Besides this, the Pareto chart (Figure 4C) shows the only significant effect is the interaction term between tetracaine and cholesterol concentration. This term is in bold in Equation 6.

$$D = -488.64 + 306.94 * TtC - 0.86 * TtC^2 + 46.69 * Co + 0.52 * Co^2 - 11.25 * TtC * Co \quad (6)$$

The results showed the influence of the concentration of cholesterol and tetracaine on diameters of liposomes produced. High concentrations of tetracaine and low concentrations of cholesterol, as well as the inverse (low concentrations of tetracaine and high concentrations of cholesterol), favor the formation of liposomes with large sizes. Probably, this is due to the competition of these molecules (tetracaine or cholesterol) in the lipid bilayer at the time of liposomes formation. When both concentrations were increased, there was decrease in liposome diameter and decreased encapsulation efficiency.

Experimental design - liposomes prepared in lipophilic phase

Figure 5A and 5B show, respectively, the response surface and contour curve for encapsulation efficiency of liposomes prepared with tetracaine incorporated lipid film (lipid phase). Unlike the liposomes prepared with the drug incorporated during the lipid film hydration (or hydrophilic phase), there was no optimum value for efficiency, and the results varied between 5.6 and 83%.

The highest value found was obtained for the lowest values of tetracaine (1mg/mL) and cholesterol (25%, w/w)

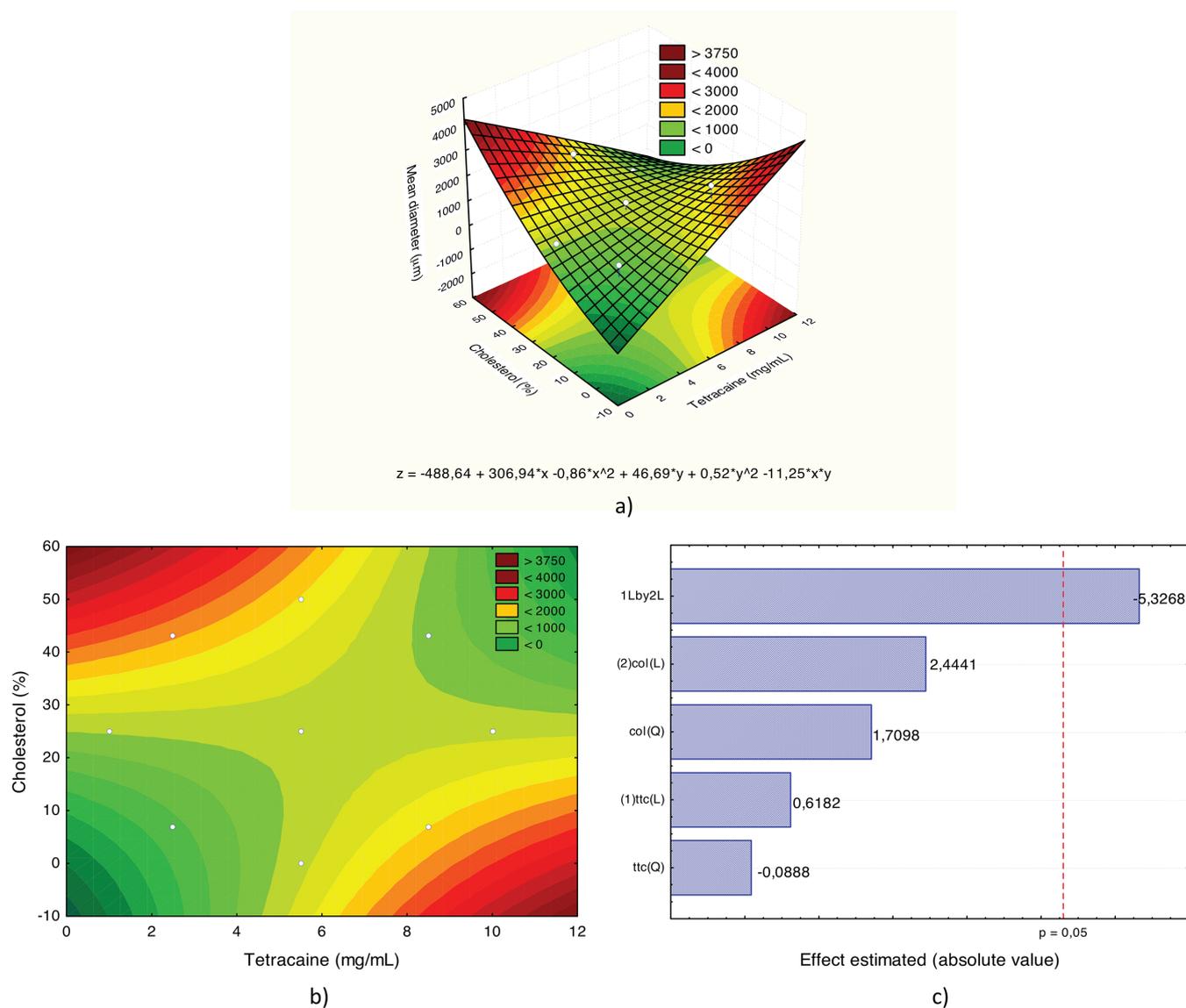


FIGURE 4 - Response surface (A), contour curve (B) and Pareto chart (C) for the mean diameter of liposomes prepared in the hydrophilic phase.

concentration into the liposomal formulation, considered the drug content compared to the lipid content is low (0.1: 75%, w/w); values correspond a low rate of encapsulation not desired.

The Pareto chart (Figure 5 C) shows that the only effect that was not significant was the quadratic term of the variable cholesterol concentration. Equation 7 shows the theoretical model predicted, the significant terms of the model are in bold.

$$E = \mathbf{50.51} - \mathbf{9.19*TTc} + \mathbf{0.98*TTc^2} + \mathbf{0.56*Co} - 0.00010*Co^2 - \mathbf{0.16*TTc*Co} \quad (7)$$

The results demonstrate that despite the tetracaine being a lipophilic local anesthetic, the better drug

encapsulation was achieved in the hydrophilic phase rather than in the lipophilic phase, as expected. Studies show that tetracaine incorporated into synthetic (phosphatidylcholine and phosphatidylserine) and biological membranes are expelled from the bilayer by pressure increase (Auger *et al.*, 1987).

Skou (1954) and Seeman (1966) showed the effect of anesthetic in lipid phases. The results presented the local anesthetics incorporation by the lipid phase causes an expansion of the surface area of monolayers (Skou, 1954) and bilayers (Seeman, 1966). This expansion is favored by the difference between the anesthetic molecule length (shorter) and phospholipids. Thus, the anesthetic would create “void volume” below its insertion point, which would be offset by a conformational change of the

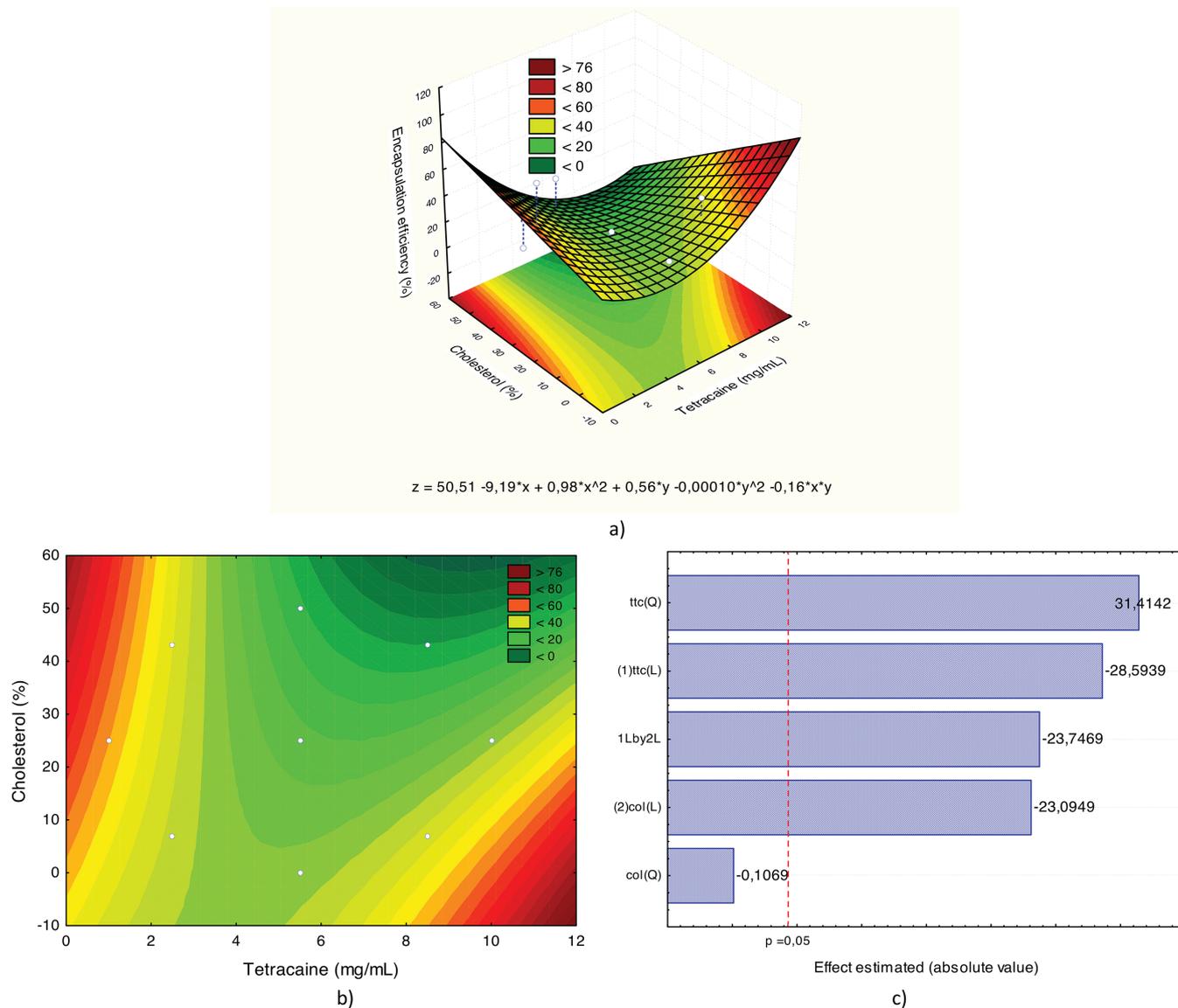


FIGURE 5 - Response surface (A), contour curve (B) and Pareto chart (C) for the encapsulation efficiency of liposomes prepared in the lipophilic phase.

adjacent lipid chains decreasing the bilayer, as well as lateral expansion (Trudell, 1977).

Experimental design with ANN

All experimental design was modeled by Equations 4, 5, 6 and 7 were also adjusted by ANN. Tables IV, V, VI, and VII show the results obtained with ANN for equations 4, 5, 6 and 7, respectively.

The ANN showed a smaller normalized error between the experimental value and the value obtained by the model (Takayama *et al.*, 2003; Subramanian, Yajnik, Murthy, 2004; Zhang *et al.*, 2010). The parameters found in the ANN are shown in Tables VIII, IX, X, and XI.

The results show that there is a substantial difference in accuracy of the ANN model compared to RSM-RCCD model (Tables IV-VII). This difference is caused due the RSM models work with regression, thus, need a specification as the linear polynomial function (second order or quadratic second order). According to Hanrahan and Lu (2006) and Ahmadi and Golian (2011), the number of terms of the polynomial is limited to points of Experimental Design. Furthermore, ANN-based models have good performance with no linear interpolation of data from different domains (Haykin, 1998).

Contrary to RSM-based methods, ANN need not specify a function, a priori, the interpolation process to act as universal approximators for any function, not only quadratic functions as the RSM-RCCD (Desai *et al.*, 2008;

TABLE IV - Comparison between experimental data and predictive models generated by RSM and ANN. Data associated with Equation 4

Assays	Diameter (nm)		
	Experimental	RSM	ANN
1	1039.0	1037.75	1039.0
2	536.2	343.08	536.2
3	210.0	560.08	210.0
4	753.8	911.89	753.8
5	1212.0	997.94	1212.0
6	698.7	756.48	698.7
7	351.7	522.79	351.7
8	915.7	586.59	915.7
9	809.4	839.21	807.72
10	806.6	839.21	807.72
11	895.8	839.21	807.72
Mean Normalized Error		0.3167	0.0093

TABLE V - Comparison between experimental data and predictive models generated by RSM and ANN. Data associated with Equation 5

Assays	Encapsulation efficiency (%)		
	Experimental	RSM	ANN
1	11.0	9.74	11.0
2	45.2	26.42	45.2
3	0.0	0.56	0.0
4	54.6	36.68	54.6
5	3.8	-4.31	3.8
6	18.1	35.29	18.1
7	21.6	19.42	21.6
8	10.7	20.17	10.7
9	28.8	32.30	33.4
10	41.4	32.30	33.4
11	30.1	32.30	33.4
Mean Normalized Error		0.5259	0.0422

Asiltürk, Çunkaş, 2011; Maran, Priya, 2015; Hanrahan, Lu, 2006).

The limitations of the techniques based on ANN are the quantity and quality of data used in its interpolation process or learning. However, the quality is more important than quantity. Some studies show that a sufficient amount of data for the ANN is equivalent to the number of experiments needed for a good regression process RSM. Other studies show that the design of experiments generates statistically

TABLE VI - Comparison between experimental data and predictive models generated by RSM and ANN. Data associated with Equation 6

Assays	Diameter (nm)		
	Experimental	RSM	ANN
1	1833	2033.11	1833.0
2	887.1	915.49	887.1
3	773.2	428.77	773.2
4	2258	1741.15	2258.0
5	1010	1028.44	1010.0
6	924.9	1174.51	924.9
7	1965	1714.27	1965.0
8	615.5	1173.52	615.5
9	968.7	1118.89	1140.3
10	996.4	1118.89	1140.3
11	1377	1118.89	1140.3
Mean Normalized Error		0.2367	0.0449

TABLE VII - Comparison between experimental data and predictive models generated by RSM and ANN. Data associated with Equation 7

Assays	Encapsulation efficiency (%)		
	Experimental	RSM	ANN
1	5.60	40.36	5.62
2	5.88	8.62	5.89
3	16.0	34.78	16.00
4	51.3	37.6	51.30
5	83.00	52.24	83.00
6	18.30	30.55	18.30
7	35.09	13.36	35.06
8	29.27	29.61	29.27
9	21.09	21.55	21.88
10	22.55	21.55	21.88
11	22.00	21.55	21.88
Mean Normalized Error		0.8972	0.0071

well-distributed data in the input field, satisfying the data quality condition (Desai *et al.*, 2008; Lou, Nakai, 2001). Thus, it can be concluded that the data used in RSM methods are sufficient for ANN training.

CONCLUSIONS

Considering the lack of information about liposomal formulations based on tetracaine, the use of experimental

TABLE VIII - Parameters found for ANN associated to data in Table IV and Equation 4

Input variables									
Bias			$x_1 = t$			$x_2 = P$			
Neuron parameters of the input layer									
$(j = 0)$			$(j = 1)$			$(j = 2)$			
$w_{1,j}^1$	-3.8716			2.3586					-3.2581
$w_{2,j}^1$	2.8818			-3.8989					-0.1912
$w_{3,j}^1$	-1.6871			2.6345					2.9997
$w_{4,j}^1$	-1.0675			0.9035					-3.8350
$w_{5,j}^1$	0.2269			3.9071					-1.0901
$w_{6,j}^1$	1.2557			2.2130					3.3801
$w_{7,j}^1$	-2.9486			-2.5385					2.9185
$w_{8,j}^1$	3.9108			2.8963					-3.1234
Neuron parameters of the hidden layer									
$w_{1,0}^2$	$w_{1,1}^2$	$w_{1,2}^2$	$w_{1,3}^2$	$w_{1,4}^2$	$w_{1,5}^2$	$w_{1,6}^2$	$w_{1,7}^2$	$w_{1,8}^2$	
-0.5588	-3.8716	2.8818	-1.6871	-1.0675	0.2269	1.2557	-2.9486	3.9108	

TABLE IX - Parameters found for ANN associated to data in Table V and Equation 5

Input variables										
Bias			$x_1 = TtC$				$x_2 = Co$			
Neuron parameters of the input layer										
$(j = 0)$			$(j = 1)$				$(j = 2)$			
$w_{1,j}^1$	4.4071					-1.9592				3.9894
$w_{2,j}^1$	3.5336					-3.0959				-3.1337
$w_{3,j}^1$	2.2213					-2.7661				-3.6703
$w_{4,j}^1$	1.5015					-0.0871				-4.4017
$w_{5,j}^1$	-0.4460					0.3748				-4.4183
$w_{6,j}^1$	0.5677					4.1302				1.3483
$w_{7,j}^1$	-1.2688					-0.8729				4.4473
$w_{8,j}^1$	-2.2461					-0.7728				-4.4547
$w_{9,j}^1$	3.3439					3.5850				-2.5824
$w_{10,j}^1$	4.6356					0.9081				4.0700
Neuron parameters of the hidden layer										
$w_{1,0}^2$	$w_{1,1}^2$	$w_{1,2}^2$	$w_{1,3}^2$	$w_{1,4}^2$	$w_{1,5}^2$	$w_{1,6}^2$	$w_{1,7}^2$	$w_{1,8}^2$	$w_{1,9}^2$	$w_{1,10}^2$
-0.183	-0.502	-0.883	0.945	0.100	-0.107	0.793	0.323	0.143	0.082	0.660

TABLE X - Parameters found for ANN associated to data in Table VI and Equation 6

Input variables											
Bias			$x_1 = Ttc$				$x_2 = Co$				
Neuron parameters of the input layer											
	($j = 0$)			($j = 1$)				($j = 2$)			
$w_{1,j}^1$	-4.3503			1.5599				4.2188			
$w_{2,j}^1$	-3.4715			3.6218				2.5979			
$w_{3,j}^1$	2.5825			-0.5429				-4.3068			
$w_{4,j}^1$	-1.4577			4.3456				0.5625			
$w_{5,j}^1$	-0.5522			3.1211				-3.1371			
$w_{6,j}^1$	-0.4053			-1.5376				-4.0681			
$w_{7,j}^1$	1.2791			2.1307				3.9591			
$w_{8,j}^1$	2.5696			3.3058				-2.806			
$w_{9,j}^1$	-3.4682			-2.8528				3.3638			
$w_{10,j}^1$	-4.2272			-0.1236				4.6009			
Neuron parameters of the hidden layer											
$w_{1,0}^2$	$w_{1,1}^2$	$w_{1,2}^2$	$w_{1,3}^2$	$w_{1,4}^2$	$w_{1,5}^2$	$w_{1,6}^2$	$w_{1,7}^2$	$w_{1,8}^2$	$w_{1,9}^2$	$w_{1,10}^2$	
-0.168	0.201	-1.494	-0.169	0.968	-0.169	0.334	0.424	0.065	0.480	0.351	

TABLE XI - Parameters found for ANN associated to data in Table VII and Equation 7

Input variables											
Bias			$x_1 = Ttc$				$x_2 = Co$				
Neuron parameters of the input layer											
	($j = 0$)			($j = 1$)				($j = 2$)			
$w_{1,j}^1$	-4.4248			1.0770				3.4484			
$w_{2,j}^1$	-2.7884			2.1154				-3.4078			
$w_{3,j}^1$	-1.7371			2.0552				3.3694			
$w_{4,j}^1$	-1.2357			3.2373				2.3255			
$w_{5,j}^1$	0.1457			-3.4256				2.0736			
$w_{6,j}^1$	-1.5048			-3.4734				2.3872			
$w_{7,j}^1$	3.2544			2.5771				-2.9282			
$w_{8,j}^1$	-3.7784			-0.6764				-4.0875			
Neuron parameters of the hidden layer											
$w_{1,0}^2$	$w_{1,1}^2$	$w_{1,2}^2$	$w_{1,3}^2$	$w_{1,4}^2$	$w_{1,5}^2$	$w_{1,6}^2$	$w_{1,7}^2$	$w_{1,8}^2$	$w_{1,9}^2$	$w_{1,10}^2$	
-0.2297	1.2364	0.3094	-0.4297	0.0452	-0.3109	1.3494	1.6949	-0.4061			

planning tool was useful in order to optimize the preparing conditions for liposomes containing this drug. It was possible to determine the optimum conditions for sonication of liposomes containing 100% lipids for obtaining LUV liposomes, these conditions were used to prepare vesicles containing the drug. According to the results of this study, it was found that the drug encapsulation efficiency was higher in the hydrophilic phase, despite tetracaine being a hydrophobic anesthetic. A comparative analysis between the RCCD-RSM and ANN techniques in process modeling and prediction surfaces was performed. The experimental results were compared using the Normalized Mean Error as used in the literature. In all cases analyzed, the ANN obtained more refined prediction errors than the RCCD-RSM technique, showing up as a viable alternative for characterizing surfaces and, in order to identify the relevant parameters which, lead to an optimization of liposomes with tetracaine for more effective local anesthesia.

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CONFLICT OF INTEREST

The author(s) declare(s) that they have no declaration of interests to disclose.

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